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# ARTICLE



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# A micro-CT based study of molar enamel thickness and its distribution pattern in Late Miocene *Lufengpithecus lufengensis* from Yunnan in Southwestern China

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## ABSTRACT

Molar enamel thickness and distribution pattern inform on the functional and dietary adaptation of extant and fossil primates. However, no systematic analysis of enamel thickness has been conducted on *Lufengpithecus*, a large-bodied fossil hominoid that lived in Southwest China during the late Miocene. In this study, we quantify two-dimensional (2D) enamel thickness and distribution of 68 lightly worn molars of *Lufengpithecus* (*L.*) *lufengensis* using micro-CT scanning data and compare it with modern humans, extant great apes, and fossil hominoids. The results indicate *L. lufengensis* has relatively thick enamel. It is slightly thicker than extant *Pongo* and comparable to some thick-enamel fossil pongines, but thinner than modern humans and most fossil hominins. The enamel distribution of *L. lufengensis* is distinctively unbalanced with relatively more enamel deposited on the cuspal region than the basal region in the molar crown, different from that found in modern humans and extant great apes. Concerning its palaeoecological and functional adaptations, we suggest that the features of thick-enamel and unbalanced distribution pattern in *L. lufengensis* is related to its adaptation to tough food and broader diets in a seasonal subtropical habitat.

## **ARTICLE HISTORY**

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#### **KEYWORDS**

Lufengpithecus lufengensis; late Miocene hominoid; micro-CT; enamel thickness; dietary adaptation

# Introduction

Lufengpithecus, a large-bodied fossil hominoid from Late Miocene Southwest China that has been unearthed with a large assemblage of isolated teeth (Xu et al. 1978; Zhang et al. 1987; Qi and Dong 2006; Xu and Lu 2008), is critical in investigating the evolution of hominoids in East Asia. It has been considered as three species: Lufengpithecus (L.) keiyuanensis, L. hudienensis, and L. lufengensis (Wu 1987; Qi et al. 2006). At present, the evolutionary history of Lufengpithecus is still unclear, although scholars published a series of study since its first discovery (e.g. Wu 1987; Kelley and Etler 1989; Qi et al. 2006; Xu and Lu 2008; Kelley and Gao 2012; Ji et al. 2013). Molar enamel thickness and distribution pattern are informative for assessing taxonomy, functional adaptation, and dietary ecology of extant and fossil primates (Molnar and Grant 1977; Kay 1981; Martin 1985; Macho and Spears 1999; Schwartz 2000; Kono et al. 2002, 2014; Smith et al. 2005, 2012a; Kono and Suwa 2008; Lucas et al. 2008; Suwa et al. 2009; Zanolli et al. 2019). It is therefore essential to investigate the enamel thickness and the dietary and ecological adaptations of Lufengpithecus.

Few previous studies have evaluated the enamel thickness of *Lufengpithecus* (Schwartz et al. 2003; Zanolli et al. 2019). Schwartz et al. (2003) analysed the ontogeny of crown formation and enamel thickness in two physical molar sections of *L. hudienensis* to compare them to *L. lufengensis* and a wide range of other hominoids, concluding that relative enamel thickness of *L. hudienensis* molar was more similar to that of *Pongo* than to any other extant ape, while *L. lufengensis* have considerably thick enamel as modern humans (Schwartz et al. 2003). Using occlusal fingerprint analysis and various morphometric approaches, Zanolli et al. (2019)

analysed hominid molars collected from the Early to Middle Pleistocene deposits of Java, Indonesia, and included *L. lufengensis* and *L. hudienensis* molars for comparison. The three-dimensional (3D) enamel distribution of *Lufengpithecus* shows relatively thick enamel at the periphery of the occlusal basin, similar to that of fossil hominid Trinil 11620 (Zanolli et al. 2019).

Although the enamel thickness and distribution of *Lufengpithecus* were preliminarily investigated, there is still no large-scale systematic analysis. In this study, we quantify the enamel thickness of *Lufengpithecus* by selecting 68 *L. lufengensis* molars from the Shihuiba site in Lufeng County, Yunnan, China (as demonstrated in Table 1), where the most abundant isolated teeth and other fossil remains of *Lufengpithecus* unearthed (Xu and Lu 2008). Micro-CT technique was used to scan the teeth and create virtual models. We aimed to analyse the 2D enamel thickness and distribution pattern of *L. lufengensis*, and compare it with modern humans, extant great apes, and other fossil hominoids. Concerning its palaeoecology and dietary adaptation, we try to provide the new dental information in understanding the ecological and functional adaptation of *Lufengpithecus*.

# **Materials and methods**

# Samples

A total of 68 isolated molars of *L. lufengensis* from Shihuiba site in Lufeng County, Yunnan, were collected, including 32 upper molars and 36 lower molars (Table 1). Sex was not incorporated into our analysis as a variable. We also collected the comparative samples, 25 molars from modern humans and 41 molars from extant great apes

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Table 1. L. lufengensis and comparative specimens in present study.

Taxon	M <sup>1</sup>	M <sup>2</sup>	M³	M <sub>1</sub>	$M_2$	$M_3$	Total	Occlusal wear <sup>a</sup>	Sources <sup>b</sup>
L. lufengensis	9	16	7	5	16	15	68	66(1), 2(2)	IVPP
Homo sapiens	2	4	3	5	7	4	25	24(1), 1(2)	IVPP
Pongo pygmaeus	2	2	2	2	2	2	12	8(1), 4(2)	MCZ (6), IVPP (6)
Pan troglodytes	1	4	2	1	3	3	14	13(1), 1(2)	AMNH (7), IVPP (7)
Gorilla gorilla	2	1	2	4	3	3	15	13(1), 2(2)	AMNH (5), IVPP (10)

<sup>a</sup>The wear stage is estimated according to (Molnar 1971). e.g. 66(1) means 66 teeth are in occlusal wear 1.

<sup>b</sup>IVPP: Institute of Vertebrate Palaeontology and Palaeoanthropology, Beijing, China MCZ: Museum of Comparative Zoology, Harvard University, Cambridge, MA, USA (www.morphosource.org)

AMNH: American Museum of Natural History, New York, New York, USA (www. morphosource.org)

e.g. AMNH (5) means five specimens are provided by AMNH.

(Table 1) from the American Museum of Natural History (AMNH), Museum of Comparative Zoology (MCZ), and Institute of Vertebrate Palaeontology and Palaeoanthropology (IVPP) (see details in Table1).

Table 1. L. lufengensis and comparative specimens in present study

# Micro-computed tomography and virtual reconstruction

The teeth provided by IVPP were scanned using the 225 kV microcomputerised tomography (developed by the Institute of High Energy Physics (IHEP), Chinese Academy of Sciences (CAS)) at the Key Laboratory of Vertebrate Evolution and Human Origins, CAS. The scanning was carried out with beam energy of 140kV and a flux of 120  $\mu$ A using a 360° rotation with a step size of 0.5°. A total of 720 projections were reconstructed into a 2048 × 2048 pixels of 1536 slices using a two-dimensional reconstruction software developed by IHEP. Isometric voxel size ranged from 10.04 to 62.70  $\mu$ m. Enamel and dentine of each specimen were digitally segmented in MIMICS 16.0 (www.materialise.com). For teeth showing occlusal wear of early phase of stage 2 (Table 1), the missing enamel was reconstructed employing unworn occlusal surface as models. Reconstructions were also made for fractures on some teeth (see details in SOM Figure 1).

#### Two-dimensional measurements of enamel thickness

Martin (1983, 1985) established widely used 2D enamel thickness measurement protocols in the ideal mesial planes. Benazzi et al. (2014) summarised the methods to identify the ideal mesial planes. In this study, we used the '2D-a' method described by Tafforeau (2004) and Benazzi et al. (2014): the mesial cusp sections (MCS) pass through two mesial dentine horn tips and are perpendicular to the best-fit plane of the cervical line (as described in Figure 1). 2D enamel thickness was measured using 3-matic 6.0 (www.materialise.com). First, we applied the 'Create Datum Plane' option in the 'Analyse' module to create the best-fit plane of the cervical line and the mesial cusps plane (Figure 1c). Second, the enamel and dentine models were cut by the mesial cusp plane through the "Cut" o'tio' in the "Design' modul' (Figure 1d). Third, we computed the 2D enamel thickness data through the "Measur" and "roper'ies" modul'.

In the 2D measurements, the variables obtained for each specimen are categorised into two groups.

Group 1: Measurements of the whole MCS (Figure 1e) (Martin 1983, 1985)

1. EA, the enamel area, in  $mm^2$ .

2. DA, the area of dentine and pulp enclosed by EDJ and straight line between the buccal and lingual cervices, in mm<sup>2</sup>.

3. EDJL, the enamel-dentine junction (EDJ) length, in mm.

4. 2DAET, the two-dimensional average enamel thickness, in mm. 2DAET is the quotient of EA and EDJL, yielding the average straight-line distance from the EDJ to the outer enamel surface. 2DAET was calculated as: 2DAET = EA/EDJL.



Figure 1. 2D enamel thickness measurements on virtual mesial cusp sections of L. lufengensis molar (M<sup>1</sup>, PA674.48). a, segmentation of enamel and dentine; b, dentine model; c and d, the mesial cusp section; e, enamel area, dentine area, and enamel-dentine junction (EDJ) length measured on the mesial cusp section; f, the cusp and basal region.

5. 2DRET, the two-dimensional relative enamel thickness, a scale-free measurement. 2DRET is 2DAET divided by the square root of dentine area, then multiplied by 100. 2DRET was calculated as:  $2DRET = 100 \times 2DAET/DA^{1/2}$ .

Group 2: Measures of the enamel distribution.

To compare enamel distribution patterns, the MCS was divided into two parts: cuspal and basal (Figure 1f). The boundary between cuspal and basal parts was set at the line passing through the lowest point of the enamel between the lingual and buccal cusp tips, parallel to the straight line between the buccal and lingual cervices (modified from Kono 2004). Average and relative thickness were calculated for each subdivision.

1. CEA and BEA, enamel area of the cuspal region and enamel area of the basal region, in  $mm^2$ .

2. CEDJL and BEDJL, EDJ length of the cuspal region, and EDJ length of the basal region, in mm.

3. CAET, the quotient of CEA and CEDJL, yielding the average straight-line distance from the EDJ to the outer enamel surface of the cusp region. CAET was calculated as: CAET = CEA/CEDJL.

4. BAET, the quotient of BEA and BEDJL, yielding the average straight-line distance from the EDJ to the outer enamel surface of the basal region. BAET was calculated as: BAET = BEA/BEDJL.

5. CRET, CRET is CAET divided by the square root of the whole dentine area (DA in Group 1), then multiplied by 100. CRET was calculated as: CRET =  $100 \times CAET/DA^{1/2}$ .

6. BRET, BRET is BAET divided by the square root of the whole dentine area (DA in Group 1), then multiplied by 100. BRET was calculated as: BRET =  $100 \times \text{BAET/DA}^{1/2}$ .

# 3 Results

# **Enamel thickness**

The results of enamel thickness measurements of *L. lufengensis* are listed in Table 2. The mean value of 2DAET is 1.08 mm with a range of 0.73–1.81 mm. The second and the third molar possess larger mean values of 2DAET than the first molar on both maxilla and the mandible, especially  $M_3$  is significantly larger than  $M_1$  in 2DAET. *L. lufengensis* 2DAET values range widely, similar to *Homo* (*H.*) *sapiens*. Nearly half of their ranges overlap. The *L. lufengensis* data almost completely overlap with the ranges of extant apes (Figure 2).

For 2DRET, values range from 11.36-28.40 (average = 17.95), and it increases from the first molar to the third moalr. Here, the overlap of *L. lufengensis* and *H. sapiens* is small in per tooth type

 Table 2. Measurements of 2D molar enamel thickness of L. lufengensis.

		Enamel area	Dentinearea	2	2DAET	
		(mm²)	(mm²)	EDJ Length (mm²)	(mm)	2DRET
Tooth	n	Mean & range	Mean & range	Mean & range	Mean & range	Mean & range
M <sup>1</sup>	9	16.21	33.02	17.30	0.93	16.21
		(11.27–22.60)	(24.71–39.78)	(14.56–19.11)	(0.76–1.18)	(12.89–20.38)
M <sup>2</sup>	16	24.28	46.47	20.64	1.17	17.32
		(17.71–39.90)	(31.86–64.74)	(17.28–25.40)	(0.94–1.81)	(13.15–23.46)
M <sup>3</sup>	7	18.95	34.42	17.11	1.11	19.43
		(13.55–24.25)	(23.10–56.15)	(13.96–21.43)	(0.96–1.37)	(12.87-24.20)
M <sub>1</sub>	5	14.01	35.78	17.59	0.79	13.44
		(11.60–16.73)	(25.82–48.82)	(15.24–19.81)	(0.73–0.84)	(12.09–15.25)
M <sub>2</sub>	16	20.87	40.51	18.94	1.10	17.73
		(16.34–27.16)	(27.77–60.04)	(15.84–22.82)	(0.88–1.38)	(11.36–24.35)
M <sub>3</sub>	15	17.98	29.39	16.14	1.11	20.70
		(13.86–21.68)	(25.01–37.53)	(14.78–18.34)	(0.89–1.44)	(16.73-28.40)
Mean	68	19.72	37.49	18.22	1.08	17.95
		(11.27–39.90)	(23.10-64.74)	(13.93–25.40)	(0.73–1.81)	(11.36-28.40)



Figure 2. Comparison of the 2D average enamel thickness (2DAET). Standard box and whisker plots reveal the interquartile range (boxes: 25%-75% percentiles of the data), 1.5 interquartile ranges (whiskers) and the median values (black line in a box). Outliers more than 1.5 interquartile from the box are indicated by circles.

(Figure 3). *Pongo* 2DRET values fall in the middle range of *L. lufengensis* values, while *Pan* and *Gorilla* fall in the lower end of the *L. lufengensis* values (Figure 3).

Table 3 lists the 2D relative enamel thickness (2DRET) of *L. lufengensis*, and extant and fossil hominoids. In general, *L. lufengensis* has thinner enamel than most other hominins, comparable to thick-enamel members of pongines. The 2DRET results indicate that most extant and fossil hominins have 'thick' (e.g. *H. sapiens, Australopithecus afarensis*, African *H. erectus*) or even 'hyper thick' (*Paranthropus robustus*) enamel (Table 3). *L. lufengensis* is similar to Asian *H. erectus* in  $M^1$  and  $M^2$ . The range of extant and fossil pongines in Asia is wide, from intermediate/thin (extant *Pongo abelli*) to thick (*Sivapithecus, Khoratpithecus*, and *Gigantopithecus*). The mean value of *L. lufengensis* is slightly thicker than that of extant *Pongo* in per tooth type (Table 3). However, there are significant overlaps between *L. lufengensis* and living and fossil orangutans in 2DRET (Smith et al. 2011, 2012b, 2018).

Previous results for Lufengpithecus fall into the 2DRET range determined in this study (Schwartz et al. 2003; Zanolli et al. 2019). The 2DRET of L. lufengensis based on physical sections (Schwartz et al. 2003) was 24.2 (M<sub>2</sub>), which is as thick as that found for H. sapiens and early hominins. It is nearly equal to the maximum 2DRET for M<sub>2</sub> found in this study (24.35). While L. hudienensis show an 'intermediate/thin' enamel thickness (2DRET =  $14.1, M_1$ ) (Schwartz et al. 2003). It is close to the mean value of 2DRET for M<sub>1</sub> of L. lufengensis in this study (13.44). Another study of Lufengpithecus based on CT scanning showed that the mean value of 3D relative enamel thickness (3DRET) in 7 molars of L. lufengensis and L. hudienensis was 15.66 (all molar positions were uncertain) (Zanolli et al. 2019), which is slightly thinner than the mean value in this study (17.95). The present study indicates the sample size and the tooth type are effective for the results of enamel thickness comparison.

# Enamel distribution pattern

The 2D enamel thickness for the whole enamel area (EA, 2DAET and 2DRET), cuspal region (CEA, CAET, and CRET), and basal region (BEA, BAET, and BRET) are provided in Table 4. The data for each species are separated by upper and low molars, although differences in molar enamel thickness can also exist within the upper and lower rows (see Table 2).

The results of enamel thickness distribution are shown in three ways: 1) comparisons of cuspal enamel area (CEA) and basal enamel area (BEA); 2) comparisons of CAET and BAET; 3) comparisons of 2DRET, CRET, and BRET among taxa.

The proportions of cuspal and basal enamel areas in upper and lower molars are shown in Figures 4 and 5. Cuspal enamel area is larger than basal enamel area, especially in *L. lufengensis* and *H. sapiens* (Table 4, Figures 4 and 5). The CEA/BEA shows the ratio of cuspal enamel area to basal enamel area. We compared CEA/BEA using a Mann Whitney U-test to test for significant differences between *L. lufengensis* and comparative specimens (Table 5). *L. lufengensis* and *H. sapiens* are significantly different in upper molars but similar in lower molars, while *L. lufengensis* and other hominoids are significantly different in both upper and lower molars (Table 5).

Cuspal enamel (CAET) is thicker than basal enamel (BAET) in all species examined (Table 4). For both upper and lower molars, *H. sapiens* shows the thickest CAET and *L. lufengensis* ranks second. In contrast, for BAET, *L. lufengensis* has the thinnest enamel. This suggests that the enamel distribution of *Lufengpithecus* molars shows relatively more enamel deposits in the cuspal than the basal region. Cuspal and basal average enamel thickness differed significantly in *H. sapiens* and *L. lufengensis* while they did not for *Pongo*, *Pan*, and *Gorilla* (Table 6).

For relative basal enamel thickness (BRET), *L. lufengensis* has the thinnest basal enamel for both upper and lower molars. In contrast, for relative cuspal enamel thickness (CRET), *L. lufengensis* ranks second (Table 4). The differences of relative enamel thickness in the whole enamel area, the cuspal region, and the basal region between *L. lufengensis* and comparative species are shown in Table 7. *L. lufengensis* shows significantly thinner enamel than *H. sapiens* for all regions. For 2DRET, *L. lufengensis* is comparable to *Pongo*, but for cuspal enamel, *L. lufengensis* is significantly thinner. *Pan* molars tend to have a thinner cuspal region, but a thicker basal enamel region than *L. lufengensis* in the whole enamel area and the cuspal enamel region, but the basal enamel of *L. lufengensis* and the basal region than *L. lufengensis*. The enamel of *Gorilla* molars is thinner than that found for *L. lufengensis* in the whole enamel area and the cuspal enamel region, but the basal enamel of *L. lufengensis* and *Gorilla* do not differ significantly from each other.

The present results reveal that *L. lufengensis* shows a distinctly unbalanced distribution pattern, different from that of modern humans and extant great apes. Relatively more enamel is deposited



Figure 3. Comparison of the 2D relative enamel thickness (2DRET). See Figure 2 for the explanation of the figure.

Taxon	M <sup>1</sup> (range)	M <sup>2</sup> (rande)	M <sup>3</sup> (rande)	M. (range)	M <sub>2</sub> (range)	M. (range)	N Mean	References	Catedorv <sup>a</sup>
Australopithecus boisei				28.74	29.42(24.82–36.65)	33.88(30.30–36.81)	8 31.01	Skinner et al. (2015)	Hyper thick
South African early <i>Homo</i>		29.5		29.0(28.7–29.3)			3 29.15	Smith et al. (2012a)	Hyper thick
Paranthropus robustus	28.82(28.61-29.03)		25.81(22.49–30.31)	28.24(24.95-30.47)	35.68		9 28.38	Olejniczak et al. (2008b)	Hyper thick
Ouranopithecus turkae					27.3		1 27.3	GüLEç et al. (2007)	Hyper thick
Australopithecus aethiopicu					26.10(25.28-26.92)	25.80	3 26.00	Skinner et al. (2015)	Thick
Ouranopithecus macedoniensis						25.5	1 25.5	Smith et al. (2004)	Thick
Homo sapiens	22.92(19.13-26.72)	22.53(19.83-26.71)	29.13(27.05–30.14)	21.95(19.13-22.96)	23.15(19.88–26.63)	28.69(25.35-33.67)	25 24.40	This study	Thick
Homo sapiens	18.7(14.0–23.9)	21.4(16.5–28.0)	21.8(17.0–30.0)	17.0(11.8–22.6)	20.5(14.8–27.7)	21.7(17.2–31.8)	271 20.04	Smith et al. (2006)	Thick
								Smith et al. (2012a)	
Australopithecus afarensis				19.49(18.37–20.61)	24.82(21.73–28.44)	24.45(24.08–24.82)	8 23.39	Skinner et al. (2015)	Thick
Afropithecus turkanensis					22.91(19.88–25.96)	24.4	4 23.28	Smith et al. (2003)	Thick
								MacLatchy et al. (2019)	
Africa Homo erectus				19.59	24.26(22.67–25.13)	23.03(20.17–25.89)	6 23.07	Skinner et al. (2015)	Thick
Australopithecus africanus	22.54		25.60	21.73(15.69–27.76)	23.93(16.54–31.32)	21.89(17.91–26.81)	9 22.79	Olejniczak et al. (2008b)	Thick
Gigantopithecus blacki	19.88(14.20-23.97)	22.45(16.76-31.02)		21.49(14.76–27.13)	25.63(22.74–30.99)		18 21.31	Zhang and Zhao (2013)	Thick
Australopithecus anamensis				21.40(16.56-25.71)	17.97(16.71–19.16)	21.27(18.87–22.65)	13 20.31	Skinner et al. (2015)	Thick
East African early <i>Homo</i>	15.63	18.67		21.50	26.18		4 20.50	Smith et al. (2012a)	Thick
Griphopithecus sp.		19.0(17.8–20.2)		17.2	18.4(16.5–20.7)	22.0(20.9–23.0)	8 19.3	Smith et al. (2003)	Thick
Sivapithecus sivalensis	17.2(16.3–18.2)					20.8	3 19.2	Martin (1985)	Thick
								Smith et al. (2006)	
Sivapithecus parvada					18.9		1 18.9	Mahoney et al. (2007)	Thick
Fossil Homo sapiens	18.2(15.7–20.9)	19.8		18.0(15.2–23.3)	18.3(16.3–20.2)	20.5(19.2–21.9)	17 18.52	Smith et al. (2012a)	Thick
Asian Homo erectus	15.8(15.3–16.3)	19.4	20.2(18.7–22.5)				6 18.48	Smith et al. (2012a)	Thick
Equatorius africanus	17.7	20.4		17.1(15.2–19.0)	17.6	17.3	6 17.87	Smith et al. (2019)	Thick
Lufengpithecus lufengensis	16.21 (12.89–20.38)	17.32 (13.15–23.46)	19.43 (12.87–24.20)	13.44 (12.09–15.25)	17.73 (11.36–24.35)	20.70 (16.73–28.40)	68 17.95	This study	Thick
Lufengpithecus lufengensis					24.2(24.1–24.6)		- 24.2	Schwartz et al. (2003)	Thick
Khoratpithecus piriyai					17.6		1 17.6	Chaimanee et al. (2006)	Thick
Khoratpithecus					17.52(17.23–17.80)		17.52 <sup>b</sup>	Chaimanee et al. (2003)	Thick
chiangmuanensis						0	, ,		/
Ekembo neseloni						17.0	0./1	beynon et al. (1998)	Intermealate/ +hick
Ekembo cf. nvanzae						16.69	1 16.69	MacLatchv et al. (2019)	unck Intermediate/
~									thick
Sivapithecus indicus	16.5						1 16.5	Mahoney et al. (2007)	Intermediate/
									thick
Homo neanderthalensis	15.2(13.80–16.93)	18.1(15.65–20.85)	18.0(14.30–18.87)	15.9(13.77–20.46)	15.7(14.21–16.80)	16.6(15.28–18.34)	42 16.44	Olejniczak et al. (2008a)	Intermediate/
				ſ					tnick
Ureopitnecus pampoili	13.0			17.4	0.71	(0.01-1.01)8.01	8.CI C	zanoili et al. (2010)	intermediate/ thick
Hylobates muelleri				13.3(10.4–15.6)	14.9(13.9–15.8)	18.5(15.0–21.8)	10 15.76	Olejniczak (2006)	Intermediate/
									thick
Pongo pygmaeus	12.03(11.82–12.25)	15.33(15.12–15.54)	17.64(17.45–17.84)	14.57(14.46–14.68)	15.29(14.83–15.74)	18.62(18.13–19.10)	12 15.58	This study	Intermediate/ +hich
Fossil <i>Pongo</i> (mainland Asia)	15.04	14.26	16.29	11.29	16.07	14.61	15 15.55	Hu and Zhao (2015)	Intermediate/
									thick
Rangwapithecus gordoni						14.9	1 14.9	Smith et al. (2003)	Intermediate/ +bicl
Disus law the same statements			16.00				1100		UTIICK Latering a diata /
Plerolapithecus catalaunicus	14.51(13.99-14.05)	(0C.41–62.41)44.48	10.88				6.84 c	Alda et al. (2013)	intermediate/ thick
Anoiapithecus brevirostris	14.09(13.94–14.24)	15.16(12.97–17.31)	14.77				7 14.80	Alba et al. (2013)	Intermediate/
									thick
Rudapithecus hungaricus Pongo pygmaeus	11.3 12.3(9.0–15.1)	13.5 15.2(10.8–18.2)	17.5 16.2(10.1–22.2)	12.8(12.3–13.4) 11.8(9.7–15.8)	15.4(15.0–15.9) 14.6(11.0–19.5)	16.1 16.3(11.2–22.5)	8 14.38 75 14.28	Smith et al. (2019) Smith et al. (2012b)	Intermediate/thin Intermediate/thin
Lufengpithecus hudienensis				14.1			1 14.1	Schwartz et al. (2003)	Intermediate/thin
									(Continued)

Table 3. 2D relative enamel thickness in L. lufengensis, extant and fossil hominoids (from greatest to least).

Table 3. (Continued).									
Тахоп	M <sup>1</sup> (range)	M² (range)	M <sup>3</sup> (range)	M <sub>1</sub> (range)	M <sub>2</sub> (range)	M <sub>3</sub> (range)	N Mean	References	Category <sup>a</sup>
Pongo abelli	12.0(9.0–16.3)	14.0(11.2–19.2)	16.8(12.2–22.1)	12.1(8.6–14.7)	14.2(11.2–18.5)	14.8(9.8–19.1)	45 13.86	Smith et al. (2012b)	Intermediate/thin
Pan troglydytes	10.58	13.50(12.38-14.36)	13.98(13.38-14.58)	11.68	14.05(13.21-14.91)	14.19(15.31–13.64)	14 13.49	This study	Intermediate/thin
Pan troglydytes	10.7(8.5–13.1)	11.7(10.7–12.5)	12.0(9.0–15.0)	12.5(10.3-14.1)	13.2(10.9–16.6)	14.1(12.4–16.6)	- 72	Smith et al. (2005)	Intermediate/thin
								MacLatchy et al. (2019)	
Pan paniscus						13.6	1 13.6	Smith et al. (2003)	Intermediate/thin
Hispanopithecus laietanus	13.1(12.7–13.4)	14.3					3 13.47	Andrews et al. (1991)	Intermediate/thin
•								Smith et al. (2019)	
Proconsul major				12.0(11.4–12.6)	14.7(12.3–17.2)	13.7	5 13.43	MacLatchy et al. (2019)	Intermediate/thin
								Smith et al. (2003)	
cf. Khoratpithecus sp.		13.35(13.11-13.58)					2 13.35	Chaimanee et al. (2019)	Intermediate/thin
Dryopithecus fontani		12.48	12.95				2 12.72	Alba et al. (2013)	Intermediate/thin
Symphalangus syndactylus				10.8(9.02-12.65)	13.6(11.8–15.7)	12.7(10.5–15.6)	17 12.58	Olejniczak (2006)	Intermediate/thin
Nyanzapithecus alesi	12.2						1 12.2	Nengo et al. (2017)	Intermediate/thin
Gorilla gorilla	11.82(11.69–11.96)	12.08	15.83(15.75-15.91)	9.77(9.27-10.32)	12.11(11.10-12.93)	13.30(11.07-14.47)	15 12.18	This study	Intermediate/thin
Gorilla gorilla	9.3(7.0–10.2)	10.7(8.9–13.0)	10.8(9.6–12.4)	9.1(7.8–12.6)	11.4(8.6–15.2)	12.7(11.1–15.1)	- 38	Smith et al. (2005)	Thin
								MacLatchy et al. (2019)	
Morotopithecus bishopi			12.9		11.1		2 11.98	MacLatchy et al. (2019)	Intermediate/thin
Hylobates lar						11.0	1 11.0	Martin (1983)	Thin
Proconsul africanus	8.5						1 8.5	Andrews et al. (1991)	
<sup>a</sup> Enamel thickness categories an greater than that of 'thick' be	e named according to 2 longs to 'hyper thick' (0	2DRET (Martin 1985): t Grine and Martin 1988	hick (17.49< RET≤26.2 ).	:0), intermediate/thic	:k (14.64< RET≤17.49),	intermediate/thin (11.	30< RET≤14	.64), thin (8.90< RET≤11.30)	. The value of 2DRET

<sup>b</sup> predet that the total of the mean vertices combined; male only is 17.23 and female only is 17.80 (Chaimanee et al. 2003).

Table 4. Enamel thickness of the whole enamel area, cuspal region, and basal region for upper (UM) and lower molars (LM).

				۷	Vhole ena	amel ar	ea				Cuspal	region					Basal	region		
Tooth	Taxon	n	EA	SD	2DAET	SD	2DRET	SD	CEA	SD	CAET	SD	CRET	SD	BEA	SD	BAET	SD	BRET	SD
UM	Ното	9	26.14	3.57	1.44	0.18	24.82	4.06	22.01	2.91	1.86	0.27	31.95	5.75	4.13	1.23	0.64	0.10	10.91	1.07
	Lufengpithecus	32	20.85	5.92	1.09	0.22	17.47	3.10	16.23	4.86	1.43	0.35	22.94	4.88	4.66	1.54	0.60	0.11	9.59	1.38
	Pongo	6	21.31	4.32	1.03	0.17	15.00	2.31	15.05	4.55	1.17	0.27	16.94	3.64	6.26	0.43	0.80	0.02	11.59	0.53
	Pan	7	17.60	2.84	0.86	0.10	13.22	1.31	11.61	2.62	0.93	0.16	14.32	2.18	5.99	0.74	0.74	0.05	11.45	0.97
	Gorilla	5	28.90	3.95	1.11	0.15	13.48	1.93	20.36	3.82	1.25	0.21	15.24	2.68	8.54	0.61	0.86	0.07	10.55	1.10
LM	Ното	16	22.23	3.64	1.32	0.18	24.16	3.65	17.52	2.53	1.72	0.30	31.39	5.70	4.71	1.78	0.70	0.13	12.76	1.96
	Lufengpithecus	36	18.57	3.71	1.05	0.16	17.98	3.52	14.50	3.84	1.39	0.23	23.83	4.72	3.80	0.88	0.54	0.07	9.31	1.67
	Pongo	6	21.49	1.72	1.08	0.09	16.16	1.81	15.62	1.15	1.27	0.13	19.03	2.46	5.86	0.96	0.77	0.06	11.52	1.00
	Pan	7	16.87	3.22	0.86	0.11	13.77	1.10	12.33	3.27	0.96	0.17	15.31	1.89	4.53	0.55	0.66	0.05	10.72	1.04
	Gorilla	10	27.19	6.50	0.98	0.18	11.53	1.81	19.40	5.78	1.08	0.23	12.59	2.32	8.15	1.14	0.81	0.08	9.55	0.70



Figure 4. The proportions of the cuspal and basal enamel area in the upper molars.





on the cuspal region (or upper part) than the basal region (or lower part) of the molar crown. The main reason seems to be that *L. lufengensis* shows a relatively thinner and longer enamel distribution pattern near the cervical line in the basal enamel region, compared with *H. sapiens* and extant great apes (Figure 6, also see BAET and BRET in Table 4).

The mesial cusp sections are derived from microtomography. The left side of each section is lingual, and the right side is buccal. The long red lines pass through the lowest point of the enamel between the lingual and buccal cusp tips. The short red lines are cervical lines, the straight line connecting the buccal and lingual cervices. These two types of red lines are parallel, dividing the

Table 5. Results of the Mann-Whitney U-test for differences in CEA/BEA between *L. lufengensis* and comparative specimens.

	1 1			
Group 1	Group 2		UM	LM
Lufengpithecus	Ното	Р	0.000	0.685
		Z	3.497	0.405
	Pongo	Р	0.016	0.014
	-	Z	-2.363	-2.413
	Pan	Р	0.000	0.020
		Z	-3.807	-2.296
	Gorilla	Р	0.002	0.000
		Z	-2.889	-3.579

The bold number indicates that the group comparison is significant (P < 0.05).

Table 6. Results of the Mann-Whitney U-test for differences between CAET and BAET.

Tooth		Lufengpithecus	Ното	Pongo	Pan	Gorilla
UM	Р	0.000	0.000	0.026	0.026	0.008
	Z	-6.877	-3.578	-2.181	-2.239	-2.611
LM	Р	0.000	0.000	0.002	0.007	0.003
	Ζ	-6.983	-4.824	-2.882	-2.619	-2.873

The bold number indicates that the group comparison is significant (P < 0.001).

Table 7. Results of the Mann-Whitney U-test for differences in 2DRET, CRET, and BRET between *L. lufengensis* and comparative specimens.

				UM			LM	
Group 1	Group	2	2DRET	CRET	BRET	2DRET	CRET	BRET
Lufengpithecus	Ното	Ρ	0.000	0.000	0.007	0.000	0.000	0.000
		Ζ	3.811	3.402	2.630	4.413	3.987	4.626
	Pongo	Ρ	0.106	0.010	0.001	0.171	0.014	0.008
	5	Ζ	-1.641	-2.522	3.003	-1.401	-2.413	2.569
	Pan	Ρ	0.000	0.000	0.001	0.004	0.000	0.030
		Ζ	-3.294	-3.916	3.111	-2.794	-3.720	2.154
	Gorilla	Ρ	0.007	0.001	0.140	0.000	0.000	0.470
		Ζ	-2.577	-3.021	1.511	-4.255	-4.628	0.747

The bold number indicates that the group comparison is significant (P < 0.05).

mesial cusp section into two parts: cuspal and basal. The missing enamel over cusp tips was reconstructed (the green area in 'h').

a:  $M^1$ , L. lufengensis; b:  $M^2$ , H. sapiens; c:  $M^2$ , Pongo pygmaeus; d:  $M^2$ , Pan troglodytes; e:  $M^3$ , Gorilla gorilla

f:  $M_2$ , L. lufengensis; g:  $M_1$ , H. sapiens; h:  $M_2$ , Pongo pygmaeus; i:  $M_2$ , Pan troglodytes; j:  $M_2$ , Gorilla gorilla

# Discussion

Enamel thickness has been widely used to interpret functional and dietary adaptation of primates (Molnar and Gantt 1977; Kay 1981; Dumont 1995; Shimizu 2002; Vogel et al. 2008; Kono et al. 2014). Molnar and Gantt (1977) first discussed the functional significance of enamel based on linear measurements of enamel thickness of extant great apes and modern human molars. They concluded that human teeth with thick enamel and low cusps were able to adapt to a crushing-grinding function for a long period. Kay (1981) established an association between thick enamel and diets of hard fruit. seeds, and nuts, such that thin enamel would be consistent with a diet of less abrasive and soft food. Pongo has relatively thick enamel, which might be an adaptation to eating very hard fruits, while Gorilla has relatively thin enamel and well-developed shearing crests, which are likely to be adaptations to leaf-eating (Kay 1981). Moreover, Kono (2004) explored the 3D enamel distribution pattern in 74 extant great apes and modern human molars. The examination supported the view that the functional side of the molars had thicker enamel than the opposite side (e.g. Macho and Berner 1993; Schwartz 2000; Kono et al. 2002). However, it should be noted that there is no specific value of enamel thickness that separates hard- and soft-object feeders. Dietary inferences based on the thickness of enamel should be made within an appropriate context (Dumont 1995). L. lufengensis shows a wide range of 2DRET, thinner than most of hominins, but similar to Asian *H. erectus* in  $M^1$  and  $M^2$  (Table 4). When compared with members of pongines, L. lufengensis is comparable to thick-enamel fossil pongines, such as Sivapithecus and Khoratpithecus, and they overlap with a wide range of living and fossil orangutans (Smith et al. 2011, 2012b, 2018). However, the comparison of enamel thickness distribution demonstrates that the cuspal enamel of L. lufengensis is significantly thicker than that of Pongo. This indicates that L. lufengensis is different from Pongo in the functional adaptation to its diet.

The unbalanced enamel thickness distribution associated with the abrasive diet may be related to the environment in which *L. lufengensis* lived. Fossils of *L. lufengensis* with a flora and a mammalian fauna were unearthed from beds 2–6 in section D at the Shihuiba site (Qi 1993; Xu and Lu 2008). Judging from the deposits, floral and faunal communities, Qi (1993) proposed that these hominoids lived in a woodland environment with bushes and grass at the edges. The spore-pollen assemblage showed the predominant fruits such as *Myrica* and some nuts including



Figure 6. Virtual mesial sections of molars of L. lufengensis and comparative specimens.

Corylus, Carya, and Juglans (Sun and Wu 1980), which were edible for Lufengpithecus (Xu and Lu 2008). Chang et al. (2015) analysed the pollen data from Shuitangba section, another site bearing fossil remains of L. lufengensis (Ji et al. 2013). They concluded that when the hominoid lived, evergreen broad-leaved forests with evergreen Quercus were predominant, while grasses including Poaceae began to expand, and simultaneously conifers decreased, indicating a tropical or subtropical climate. But the amounts and the kinds of spores and pollens varied through the different beds, indicating the climate changed from warm-humid to warm-drier then coolerhumid. This produced a shift from the woodland type to an environment with deciduous arboreal and bush vegetation (Sun and Wu 1980; Qi 1993; ). Furthermore, 85% of Lufengpithecus permanent teeth display a semi-annual pattern of linear enamel hypoplasia (LEH) that might be caused by the twice-yearly nutritional stress (Zhao 2004; Wang and Zhao 2015). Previous investigation of modern primate dietary ecology concluded that most primates preferred ripe fruit when available but consume 'fallback' foods when preferred foods are unavailable (Wrangham 1980; Chapman and Chapman 1990; Marshall and Wrangham 2007). For example, Lophocebus albigena have thick enamel and are commonly considered a hard-object consumer, but they consumed hard foods (e.g. seeds and bark) as 'fallbacks' when the softer foods (e.g. highquality rape fruits) were unavailable during critical periods (Lambert et al. 2004). It noted to us that what primates eat during their lifetimes may differ from what they are adapted to eating (Guatelli-Steinberg 2018). Although we demonstrated that L. lufengensis molars have more enamel deposited on the cuspal region of the dental crowns than that of extant great apes, the occlusal surface with fine wrinkles are always flat, smooth, or heavily worn (Xu and Lu 2008). This may imply that L. lufengensis have taken fallback food - perhaps nuts, seeds, or even bark - when their preferred food was unavailable due to seasonal climate change.

In addition, the postcranium bones of *Lufengpithecus* reveal that it could be well adapted to arboreal and terrestrial substrates simultaneously (Xu and Lu 1986, 2008; Lin et al. 1987). The scapula, the clavicle, and the proximal phalanges display some specialised suspensory features (pear-shaped glenoid cavity, the twisted clavicle shaft, developed fibrous flexor sheaths and moderate phalangeal curvature, etc.) as seen in the tree-dwelling orangutans (Xu and Lu 1986, 2008; Deane and Begun 2008). Compared with the upper limb bones, the lower limb bones possess more terrestrial characteristics. The proximal femur shows the ability to walk bipedally with short steps, while the first metatarsal morphology (the ellipsoid-shaped central articular surface and relatively straight shaft) is similar to that of terrestrial hominoids (Xu and Lu 2008). Thick enamel may be related to increasing degrees of abrasion in the diet related to the use of foods from terrestrial sources, and/or to the incorporation of particularly tough food such as may be present in more strongly seasonal and harsher environments (Andrews et al. 1997). Thus, thick enamel in the cuspal region, different from that of arboreal orangutan, may be helpful for L. lufengensis if they forage for diverse food in the open ground environment during the dry season.

Future studies including 3D enamel distributions, enameldentine junction (EDJ) morphology, and the growth pattern of the whole dentition would be helpful to better understand the taxonomy, functional, and dietary adaptation of *Lufengpithecus*.

# Conclusions

The present study indicates that *L. lufengensis* has relatively thick enamel. It is comparable to thick-enamel members of pongines, but

thinner than modern human and most fossil hominins. In enamel distribution, *L. lufengensis* shows a distinctly unbalanced distribution pattern, relatively with more enamel deposited on the cuspal region than the basal region in the molar crown, which is different from that found in *Pongo*, and also different from that found in modern humans and extant Africa great apes. Concerning its palaeoecological and functional adaptations, we suggest that the features of thick-enamel and unbalanced distribution pattern in *L. lufengensis* is related to its adaptation to tough food and broader its diets in a seasonal subtropical habitat.

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The authors declare that they have no conflict of interest.

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#### References

- Alba DM, Fortuny J, Pérez de Los Ríos M, Zanolli C, Almécija S, Casanovas-Vilar I, Robles JM, Moyà-Solà S. 2013. New dental remains of *Anoiapithecus* and the first appearance datum of hominoids in the Iberian Peninsula. J Hum Evol. 65(5):573–584. doi:10.1016/j.jhevol.2013.07.003.
- Andrews P, Begun DR, Zylstra M. 1997. Interrelationships between functional morphology and paleoenvironments in miocene hominoids. US: Springer.
- Andrews P, Martin L, Aiello L, Scandrett A, Andrews P, Martin L, Aiello L, Scandrett A. 1991. Hominoid dietary evolution. Philos Trans R Soc Lond B Biol Sci. 334(1270):199–209.
- Benazzi S, Panetta D, Fornai C, Toussaint M, Gruppioni G, Hublin JJ. 2014. Technical note: guidelines for the digital computation of 2D and 3D enamel thickness in hominoid teeth. Am J Phys Anthropol. 153(2):305–313. doi:10.1002/ajpa.22421.
- Beynon AD, Dean MC, Leakey MG, Reid DJ, Walker A. 1998. Comparative dental development and microstructure of *Proconsul* teeth from Rusinga Island, Kenya. J Hum Evol. 35(2):163–209. doi:10.1006/jhev.1998.0230.
- Chaimanee Y, Jolly D, Benammi M, Tafforeau P, Duzer D, Moussa I, Jaeger -J-J. 2003. A Middle Miocene hominoid from Thailand and orangutan origins. Nature. 422(6927):61–65. doi:10.1038/nature01449.
- Chaimanee Y, Lazzari V, Chaivanich K, Jaeger -J-J. 2019. First maxilla of a late Miocene hominid from Thailand and the evolution of pongine derived characters. J Hum Evol. 134:102636. doi:10.1016/j.jhevol.2019.06.007.
- Chaimanee Y, Yamee C, Tian P, Khaowiset K, Marandat B, Tafforeau P, Nemoz C, Jaeger JJ. 2006. *Khoratpithecus piriyai*, a late Miocene hominoid of Thailand. Am J Phys Anthropol. 131(3):311–323. doi:10.1002/ajpa.20437.
- Chang L, Guo Z, Deng C, Wu H, Ji X, Zhao Y, Zhang C, Ge J, Wu B, Sun L, et al. 2015. Pollen evidence of the palaeoenvironments of *Lufengpithecus lufengensis* in the Zhaotong Basin, southeastern margin of the Tibetan Plateau. Palaeogeogr Palaeoclimatol Palaeoecol. 435:95–104. doi:10.1016/j. palaeo.2015.06.007.
- Chapman CA, Chapman LJ. 1990. Dietary variability in primate populations. Primates. 31(1):121–128. doi:10.1007/BF02381035.
- Deane AS, Begun DR. 2008. Broken fingers: retesting locomotor hypotheses for fossil hominoids using fragmentary proximal phalanges and high-resolution polynomial curve fitting (HR-PCF). J Hum Evol. 55(4):691–701. doi:10.1016/ j.jhevol.2008.05.005.

- Dumont ER. 1995. Enamel thickness and dietary adaptation among extant primates and chiropterans. J Mammal. 76(4):1127-1136. doi:10.2307/1382604.
- Grine FE, Martin LB. 1988. Enamel thickness and development in *Australopithecus* and *Paranthropus*. In: Grine FE, editor. Evolutionary history of the "Robust" Australopithecines Aldine de Gruyter. New York: Aldine de Gruyter. p. 3–42.
- Guatelli-Steinberg D. 2018. Dental anthropology in the AJPA: its roots and heights. Am J Phys Anthropol. 165(4):879–892. doi:10.1002/ajpa.23352.
- GüLEç ES, Sevim A, Pehlevan C, Kaya F. 2007. A new great ape from the late Miocene of Turkey. Anthropol Sci. 115(2):153–158. doi:10.1537/ase.070501.
- Hu R, Zhao L. 2015. CT analysis on enamel thickness of pleistocene fossil Orangutan from Guangxi, South China. Acta Anthropol Sin. 34(3):13.
- Ji X, Jablonski NG, Su DF, Deng C, Flynn LJ, You Y, Kelley J. 2013. Juvenile hominoid cranium from the terminal Miocene of Yunnan, China. Chin Sci Bull. 58(31):3771–3779. doi:10.1007/s11434-013-6021-x.
- Kay RF. 1981. The nut-crackers a new theory of the adaptations of the Ramapithecinae. Am J Phys Anthropol. 55(2):141–151. doi:10.1002/ajpa.1330550202.
- Kelley J, Etler D. 1989. Hominoid dental variability and species number at the late Miocene site of Lufeng, China. Am J Primatol. 18(1):15–34. doi:10.1002/ ajp.1350180103.
- Kelley J, Gao F. 2012. Juvenile hominoid cranium from the late Miocene of southern China and hominoid diversity in Asia. Proc Natl Acad Sci. 109 (18):6882–6885. doi:10.1073/pnas.1201330109.
- Kono RT. 2004. Molar enamel thickness and distribution patterns in extant great apes and humans: new insights based on a 3-dimensional whole crown perspective. Anthropol Sci. 112(2):121–146. doi:10.1537/ase.03106.
- Kono RT, Suwa G. 2008. Enamel distribution patterns of extant human and hominoid molars: occlusal versus lateral enamel thickness. Bull Natl Mus Nat Sci Ser D. 34:1–9.
- Kono RT, Suwa G, Tanijiri T. 2002. A three-dimensional analysis of enamel distribution patterns in human permanent first molars. Arch Oral Biol. 47 (12):867–875. doi:10.1016/S0003-9969(02)00151-6.
- Kono RT, Zhang Y, Jin C, Takai M, Suwa G. 2014. A 3-dimensional assessment of molar enamel thickness and distribution pattern in *Gigantopithecus blacki*. Quatern Int. 354:46–51. doi:10.1016/j.quaint.2014.02.012.
- Lambert JE, Chapman CA, Wrangham RW, Conklin-Brittain NL. 2004. Hardness of cercopithecine foods: implications for the critical function of enamel thickness in exploiting fallback foods. Am J Phys Anthropol. 125 (4):363–368. doi:10.1002/ajpa.10403.
- Lin Y, Wang S, Guo Z, Zhang L. 1987. The first discovery of the radius of *Sivapithecus lufengensis* in China. Geol Rev. 33(1):1-4.
- Lucas PW, Constantino PJ, Wood BA. 2008. Inferences regarding the diet of extinct hominins: structural and functional trends in dental and mandibular morphology within the hominin clade. J Anat. 212:4. doi:10.1111/j.1469-7580.2008.00877.x.
- Macho GA, Berner ME. 1993. Enamel thickness of human maxillary molars reconsidered. Am J Phys Anthropol. 92(2):189–200. doi:10.1002/ajpa.1330920208.
- Macho GA, Spears IR. 1999. Effects of loading on the biochemical behavior of molars of *Homo, Pan*, and *Pongo*. Am J Phys Anthropol. 109(2):211–227. doi:10.1002/(SICI)1096-8644(199906)109:2<211::AID-AJPA6>3.0.CO;2-B.
- MacLatchy L, Rossie J, Houssaye A, Olejniczak AJ, Smith TM. 2019. New hominoid fossils from Moroto II, Uganda and their bearing on the taxonomic and adaptive status of Morotopithecus bishopi. J Hum Evol. 132:227–246. doi:10.1016/j.jhevol.2019.03.008.
- Mahoney P, Smith TM, Schwartz GT, Dean C, Kelley J. 2007. Molar crown formation in the Late Miocene Asian hominoids, Sivapithecus parvada and Sivapithecus indicus. J Hum Evol. 53(1):61–68. doi:10.1016/j. jhevol.2007.01.007.
- Marshall AJ, Wrangham RW. 2007. Evolutionary Consequences of Fallback Foods. Int J Primatol. 28(6):1219. doi:10.1007/s10764-007-9218-5.
- Martin L. 1985. Significance of enamel thickness in hominoid evolution. Nature. 314(6008):260–263. doi:10.1038/314260a0.
- Martin LB 1983. The relationships of the later Miocene Hominoidea. Ph D dissertation. University College London (University of London).
- Molnar S. 1971. Human tooth wear, tooth function and cultural variability. Am J Phys Anthropol. 34(2):175–189.
- Molnar S, Gantt DG. 1977. Functional implications of primate enamel thickness. Am J Phys Anthropol. 46(3):447–454. doi:10.1002/ajpa.1330460310.
- Nengo IO, Tafforeau P, Gilbert CC, Fleagle JG, Miller ER, Feibel CS, Fox DL, Feinberg J, Pugh KD, Berruyer C. 2017. New infant cranium from the African Miocene sheds light on ape evolution. Nature. 548(7666):169–174. doi:10.1038/nature23456.
- Olejniczak AJ 2006. Micro-computed tomography of primate molars. Ph D dissertation. Stony Brook University.

- Olejniczak AJ, Smith TM, Feeney RNM, Macchiarelli R, Mazurier A, Bondioli L, Rosas A, Fortea J, De La Rasilla M, Garcia-Tabernero A. 2008a. Dental tissue proportions and enamel thickness in Neandertal and modern human molars. J Hum Evol. 55(1):12–23. doi:10.1016/j. jhevol.2007.11.004.
- Olejniczak AJ, Smith TM, Skinner MM, Grine FE, Feeney RNM, Thackeray JF, Hublin -J-J. 2008b. Three-dimensional molar enamel distribution and thickness in *Australopithecus* and *Paranthropus*. Biol Lett. 4(4):406–410. doi:10.1098/rsbl.2008.0223.
- Qi G. 1993. The environment and ecology of the Lufeng hominoids. J Hum Evol. 24:9.
- Qi G, Dong W. 2006. Lufengpithecus hudienensis Site. Beijing: Science Press.
- Qi G, Dong W, Liang Z, Zhao L, Gao F, Yue L, Zhang Y. 2006. Taxonomy, age and environment status of the Yuanmou hominoids. Chin Sci Bull. 51 (6):704–712. doi:10.1007/s11434-006-0704-5.
- Schwartz GT. 2000. Taxonomic and functional aspects of the patterning of enamel thickness distribution in extant large-bodied hominoids. Am J Phys Anthropol. 111(2):221–244. doi:10.1002/(SICI)1096-8644(200002) 111:2<221::AID-AJPA8>3.0.CO;2-G.
- Schwartz GT, Liu W, Zheng L. 2003. Preliminary investigation of dental microstructure in the Yuanmou hominoid (*Lufengpithecus hudienensis*), Yunnan Province, China. J Hum Evol. 44(2):189–202. doi:10.1016/S0047-2484(02) 00197-5.
- Shimizu D. 2002. Functional implications of enamel thickness in the lower molars of red colobus (*Procolobus badius*) and Japanese macaque (*Macaca fuscata*). J Hum Evol. 43(5):605–620. doi:10.1006/jhev.2002.0593.
- Skinner MM, Alemseged Z, Gaunitz C, Hublin -J-J. 2015. Enamel thickness trends in Plio-Pleistocene hominin mandibular molars. J Hum Evol. 85:35–45. doi:10.1016/j.jhevol.2015.03.012.
- Smith T, Olejniczak A, Reid D, Ferrell R, Hublin JJ. 2006. Modern human molar enamel thickness and enamel-dentine junction shape. Arch Oral Biol. 51 (11):974–995. doi:10.1016/j.archoralbio.2006.04.012.
- Smith TM, Bacon AM, Demeter F, Kullmer O, Nguyen KT, Vos JD, Wang W, Zermeno JP, Zhao L. 2011. Dental tissue proportions in fossil orangutans from mainland Asia and Indonesia. Asian Aust Assoc Paleoanthopologists. 1:1.
- Smith TM, Houssaye A, Kullmer O, Le Cabec A, Olejniczak AJ, Schrenk F, De Vos J, Tafforeau P. 2018. Disentangling isolated dental remains of Asian Pleistocene hominins and pongines. PLoS One. 13(11):e0204737. doi:10.1371/journal.pone.0204737.
- Smith TM, Kupczik K, Machanda Z, Skinner MM, Zermeno JP. 2012b. Enamel thickness in Bornean and Sumatran orangutan dentitions. Am J Phys Anthropol. 147(3):417–426. doi:10.1002/ajpa.22009.
- Smith TM, Martin LB, Leakey MG. 2003. Enamel thickness, microstructure and development in *Afropithecus turkanensis*. J Hum Evol. 44(3):283–306. doi:10.1016/S0047-2484(03)00006-X.
- Smith TM, Martin LB, Reid DJ, Bonis LD, Koufos GD. 2004. An examination of dental development in *Graecopithecus freybergi* (= *Ouranopithecus macedoniensis*). J Hum Evol. 46(5):551–577. doi:10.1016/j.jhevol.2004.01.006.
- Smith TM, Olejniczak AJ, Martin LB, Reid DJ. 2005. Variation in hominoid molar enamel thickness. J Hum Evol. 48(6):575–592. doi:10.1016/j. jhevol.2005.02.004.
- Smith TM, Olejniczak AJ, Zermeno JP, Tafforeau P, Skinner MM, Hoffmann A, Radovčić J, Toussaint M, Kruszynski R, Menter C, et al. 2012a. Variation in enamel thickness within the genus *Homo*. J Hum Evol. 62(3):395–411. doi:10.1016/j.jhevol.2011.12.004
- Smith TM, Tafforeau P, Pouech J, Begun DR. 2019. Enamel thickness and dental development in *Rudapithecus hungaricus*. J Hum Evol. 136:102649. doi:10.1016/j.jhevol.2019.102649.
- Sun X, Wu Y. 1980. Paleoenvironment during the time of *Ramapithecus lufengensis*. Certebrata Palasiatica. 3:9.
- Suwa G, Kono RT, Simpson SW, Asfaw B, Lovejoy CO, White TD. 2009. Paleobiological implications of the Ardipithecus ramidus dentition. Science. 326(5949):69–99. doi:10.1126/science.1175824.
- Tafforeau P 2004. Phylogenetic and functional aspects of tooth enamel microstructure and three-dimensional structure of modern and fossil primate molars. Ph D dissertation, Universite de Montpellier II.
- Vogel ER, van Woerden JT, Lucas PW, Atmoko SSU, van Schaik CP, Dominy NJ. 2008. Functional ecology and evolution of hominoid molar enamel thickness: pan troglodytes schweinfurthii and Pongo pygmaeus wurmbii. J Hum Evol. 55(1):60-74. doi:10.1016/j. jhevol.2007.12.005.
- Wang C, Zhao L. 2015. New observations of linear enamel hypoplasia from late miocene *Lufengpithecus lufengensis* of Yunnan, South China. Acta Anthropol Sin. 34(4):544–552.
- Wrangham RW. 1980. An ecological model of female-bonded primate groups. Behaviour. 75(3/4):262–300. doi:10.1163/156853980X00447.
- Wu R. 1987. A revision of the classification of the Lufeng great apes. Acta Anthropol Sin. 6(4):265.

- Xu Q, Lu Q. 2008. *Lufengpithecus lufengensis*-an early member of Hominidae. Beijing: Science Press.
- Xu Q, Lu Q, Pan Y, Zhang X, Zheng L. 1978. Fossil mandible of the Lufeng Ramapithecus. Kexue Tongbao. 9:544-556.
- Xu WR, Lu Q. 1986. Relationship between Lufeng *Sivapithecus* and *Ramapithecus* and their phylogenetic position. Acta Anthropol Sin. 5 (1):1–33.
- Zanolli C, Dean C, Rook L, Bondioli L, Mazurier A, Macchiarelli R. 2016. Enamel thickness and enamel growth in *Oreopithecus*: combining microtomographic and histological evidence. Comptes Rendus Palevol. 15 (1):209-226. doi:10.1016/j.crpv.2015.02.001.
- Zanolli C, Kullmer O, Kelley J, Bacon A-M, Demeter F, Dumoncel J, Fiorenza L, Grine FE, Hublin -J-J, Nguyen AT, et al. 2019. Evidence for increased hominid diversity in the Early to Middle Pleistocene of Indonesia. Nat Ecol Evol. 3(5):755–764. doi:10.1038/s41559-019-0860-z
- Zhang L, Zhao L. 2013. Enamel thickness of *Gigantopithecus blacki* and its significance for dietary adaptation and phylogeny. Acta Anthropol Sin. 32 (3):365–376. doi:10.3724/SP.J.1011.2013.00365.
- Zhang X. 1987. New materials of *Ramapithecus* from Keiyuan, Yunnan. Acta Anthropol Sin. 6:82–86.
- Zhao L. 2004. Linear enamel hypoplasia of *Lufengpithecus lufengensis*. Acta Anthropol Sin. 23(2):111-118.